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(54) **RECOMBINANT MICROORGANISM HAVING AN ENHANCED ABILITY TO PRODUCE PUTRESCINE AND A METHOD FOR PRODUCING PUTRESCINE USING THE SAME**

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None
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(57) **ABSTRACT**

The present invention relates to a recombinant microorganism having enhanced ability to produce putrescine at high yield, wherein the activity of NCg10101 is weakened in a microorganism of genus *Corynebacterium* that has been modified to produce putrescine, and a method for producing putrescine using the same.

9 Claims, 1 Drawing Sheet

Figure 1

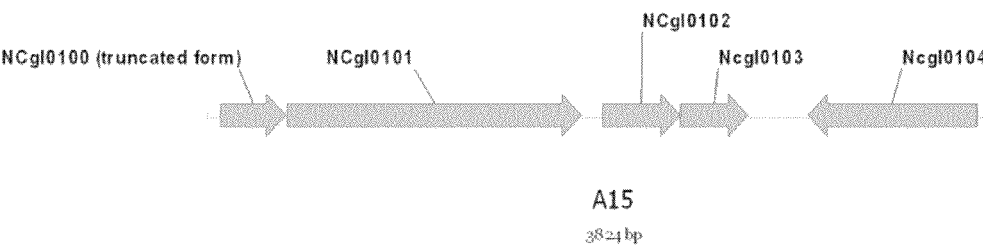
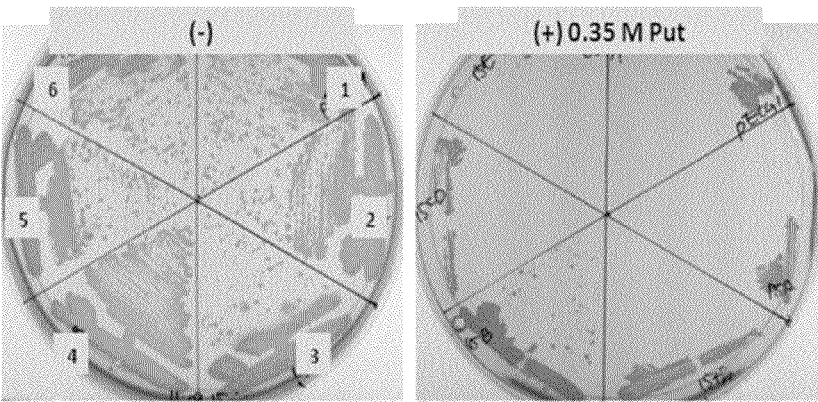


Figure 2



1

RECOMBINANT MICROORGANISM HAVING AN ENHANCED ABILITY TO PRODUCE PUTRESCINE AND A METHOD FOR PRODUCING PUTRESCINE USING THE SAME

TECHNICAL FIELD

The present invention relates to recombinant microorganisms having an enhanced ability to produce putrescine and a method for producing putrescine using the same.

BACKGROUND ART

Putrescine (or 1,4-butanediamine) is a type of polyamine, such as spermidine and spermine, and is found in gram-negative bacteria and fungi. Since putrescine is present in a wide range of concentrations in various species, it is expected to play an important role in the metabolism of microorganisms. Putrescine is commonly produced by chemical synthesis through acrylonitrile and succinonitrile from propylene. The chemical synthesis uses the substances derived from petrochemicals as starting materials and uses toxic chemicals, and thus it is not environment-friendly and has a problem of oil depletion.

In order to resolve these problems, there has been much research on developing a method for biosynthesis of putrescine by using microorganisms, that is more environment-friendly and reduces energy consumption. According to current knowledge, putrescine can be biosynthesized through two pathways. In one pathway, ornithine is produced from glutamate and the ornithine is decarboxylated to synthesize putrescine. In the other pathway, arginine is synthesized from the ornithine, agmatine is produced from the arginine, and then putrescine is synthesized from the agmatine. In addition, there are other methods for synthesizing putrescine by using a target microorganism which is transformed with the enzymes involved in the known synthetic pathways of putrescine. For example, WO09/125924 discloses a method for producing putrescine at high yield by inactivating the pathway involved in the decomposition and utilization of putrescine in *E. coli*, by inactivating the pathway in which ornithine, a precursor of putrescine, is converted to arginine, and by enhancing the biosynthetic pathway of ornithine. An article published in 2010 discloses a method for producing putrescine at high concentration by introducing and enhancing the protein that converts ornithine to putrescine into *Corynebacterium* strains which are not capable of producing putrescine. In addition it discloses a method for producing putrescine from arginine by introducing *E. coli*-derived arginine decarboxylase and agmatinase into the strains. In this regard, the ornithine pathway produced about 50 times higher amount of putrescine than the arginine pathway (Schneider et al., Appl. Microbiol. Biotechnol. 88:4, 859-868, 2010).

DISCLOSURE

Technical Problems

In this background, the present inventors identified that putrescine can be produced at high yield in a microorganism of genus *Corynebacterium* by weakening or removing the activity of NCg10101 protein (SEQ ID NOS: 17 or 19), thereby completing the present invention.

Technical Solution

One objective of the present invention is to provide a recombinant microorganism of genus *Corynebacterium*

2

capable of producing putrescine at high yield, which is modified to have the weakened NCg10101 (SEQ ID NOS: 17 or 19) activity, compared to the endogenous activity thereof.

Another objective of the present invention is to provide a method for producing putrescine using the microorganism.

Advantageous Effect

When the microorganism of genus *Corynebacterium* having an improved ability to produce putrescine of the present invention is used for the production of putrescine, it is modified to weaken NCg10101 (SEQ ID NOS: 17 or 19) activity compared to the endogenous activity thereof, and therefore, it can be produce putrescine at high yield. Accordingly, the microorganism can be widely used for the more effective production of putrescine.

DESCRIPTION OF FIGURES

FIG. 1 represents a schematic diagram showing the relative positions of genes encoding NCg10100 (SEQ ID NO: 27), NCg10101 (SEQ ID NOS: 17 or 19), NCg10102 (SEQ ID NO: 29), NCg10103 (SEQ ID NO: 30), and NCg10104 (SEQ ID NO: 31), which are on the chromosome of the wild type *Corynebacterium glutamicum* ATCC 13032 strain; and

FIG. 2 represents the test result of growth comparison between the recombinant strains prepared in the present invention, in which 1, 2, 3, 4, 5 and 6 are strains prepared by introducing pHCI39T, pHCI39T-P(CJ7)-NCg10100, pHCI39T-P(CJ7)-tNCg10100, pHCI39T-P(CJ7)-NCg10101, pHCI39T-P(CJ7)-NCg10102-NCg10103, and pHCI39T-P(CJ7)-NCg10104 into KCCM11138P, respectively.

BEST MODE

In one aspect to achieve the above objectives, the present invention provides a recombinant microorganism of genus *Corynebacterium* having an enhanced ability to produce putrescine, which is modified by weakening or removing the activity of NCg10101 protein having an amino acid sequence represented by SEQ ID NO. 17 or SEQ ID NO. 19, compared to the endogenous activity thereof.

As used herein, the term "NCg10101" means a protein showing the activity of a metal-dependent enzyme, which is expressed in *Corynebacterium glutamicum* (SEQ ID NOS: 17 or 19), and whose function is not yet fully known. It includes a metal binding domain of peptidase M20 family or aminobenzoyl-glutamate utilization protein (AbgB). The AbgB of *E. coli* constitutes aminobenzoyl-glutamate hydrolase with AbgA to hydrolyze aminobenzoyl-glutamate to aminobenzoate and glutamate. The aminobenzoate is known to be used as a precursor for folate synthesis, but its relationship with putrescine productivity has not been known.

NCg10101 protein of the present invention may comprise the amino acid sequence represented by SEQ ID NO: 17 or SEQ ID NO: 19. However, it is not limited thereto, because there may be the difference in the amino acid sequence of the protein depending on the microbial species or strains. In other words, it can be a mutant protein or artificial variant with an amino acid sequence comprising substitution, deletion, insertion, or addition of one or several amino acids at one or more locations of the amino acid sequence represented by SEQ ID NO: 17 or SEQ ID NO: 19, as long as it can help increase the ability to produce putrescine by weakening the activity of the protein. Herein, "several" may differ depending on the location or type in the three-dimensional structure of amino acid

residues of the protein, but specifically means 2 to 20, specifically 2 to 10, and more specifically 2 to 5. In addition, the substitution, deletion, insertion, addition or inversion of the amino acid includes those caused by artificial variants or natural mutation, based on the difference in the individual or species of microorganism.

The polynucleotide encoding the amino acid sequence of the present invention may comprise the polynucleotide sequence encoding the protein having amino acid sequence represented by SEQ ID NO: 17 or SEQ ID NO: 19, or the amino acid sequence of 80% or more, specifically 90% or more, more specifically 95% or more, and particularly specifically 97% or more homology with the same, as long as it has similar activity as the NCg10101 protein. The most specifically, it may be the polynucleotide sequence represented by SEQ ID NO: 16 or SEQ ID NO: 18.

The term "homology" refers to the identity between two amino acid sequences and may be determined by the well known method to those skilled in the art, using BLAST 2.0 to compute the parameter such as score, identity and similarity.

In addition, the polynucleotide sequence encoding the polypeptide with the amino acid sequence of NCg10101 (SEQ ID NOS: 17 or 19) of the present invention can be hybridized with the polynucleotide of SEQ ID. NO: 16 or the probe prepared from the same under 'stringent conditions', and may be a modified polynucleotide sequence encoding the NCg10101 protein (SEQ ID NOS: 17 or 19) which normally functions. As used herein, "stringent conditions" refer to conditions which allow the specific hybridization between the polynucleotide, and are described specifically, for example, in Molecular Cloning (A Laboratory Manual, J. Sambrook et al., Editors, 2nd Edition, Cold Spring Harbor Laboratory press, Cold Spring Harbor, N. Y., 1989) or Current Protocols in Molecular Biology (F. M. Ausubel et al., Editors, John Wiley & Sons, Inc., New York). For example, the hybridization is carried out in the hybridization buffer of 65° C. (3.5× SSC, 0.02% Ficoll, 0.02% polyvinylpyrrolidone, 0.02% bovine serum albumin, 2.5 mM NaH₂PO₄ (pH 7), 0.5% SDS, 2 mM EDTA). SSC is 0.15 M sodium chloride/0.15 M sodium citrate of pH 7. After hybridization, the membrane to which DNA is transferred is rinsed with 2×SSC at room temperature and then rinsed again with 0.1 to 0.5×SSC/0.1×SDS at a temperature of 68° C.

The activity of NCg10101 protein (SEQ ID NOS: 17 or 19) in the present invention can be weakened by 1) a partial or whole deletion of a polynucleotide encoding the protein, 2) modifying an expression regulatory sequence to reduce the expression of the polynucleotide, 3) a modification of the polynucleotide sequence on chromosome or 4) a combination thereof.

In the above, a partial or whole deletion of a polynucleotide encoding the protein can be performed by substituting the polynucleotide encoding an endogenous target protein in the chromosome to a marker gene or a polynucleotide which partial nucleotide sequence was deleted, with a vector for chromosomal gene insertion. The length of the "partial" deletion depends on the type of polynucleotide, but is specifically 2 bp to 300 bp, more specifically 2 bp to 100 bp, and further more specifically 1 bp to 5 bp.

Also, to decrease the polynucleotide expression, an expression regulatory sequence may be modified by inducing mutations in the expression regulatory sequence through deletion, insertion, conservative or non-conservative substitution of nucleotide sequence or a combination thereof to further weaken the activity of the expression regulatory sequence, or by replacing the expression regulatory sequence with the sequence having weaker activity. The expression regulatory

sequence may include a sequence encoding promoter, operator sequence, ribosomal binding site and the sequence controlling the termination of transcription and translation.

In addition, the polynucleotide sequence on chromosome to weaken the activity of the protein may be modified by inducing mutations in the sequence through deletion, insertion, conservative or non-conservative substitution of nucleotide sequence or a combination thereof to further weaken the activity of the sequence, or by replacing the polynucleotide sequence with the modified sequence to have weaker activity of the protein.

Meanwhile, a microorganism of genus *Corynebacterium* having enhanced ability to produce putrescine of the present invention may be further modified to weaken the activity of ornithine carbamoyltransferase (ArgF) involved in the synthesis of arginine from ornithine and the activity of protein (NCg11221; SEQ ID NO: 21) involved in exporting glutamate, compared to the endogenous activity thereof. In addition, the microorganism of *Corynebacterium* genus may be modified by additionally introducing the activity of ornithine decarboxylase (ODC). Also, the microorganism of genus *Corynebacterium* may be further modified to enhance the activity of acetyl glutamate synthase to convert glutamate to acetyl glutamate or ornithine acetyltransferase (ArgJ) to convert acetyl ornithine to ornithine, the activity of acetyl glutamate kinase (ArgB) to convert acetyl glutamate to acetyl glutamyl phosphate, the activity of acetyl gamma glutamyl phosphate reductase (ArgC) to convert acetyl glutamyl phosphate to acetyl glutamate semialdehyde, and the activity of acetyl ornithine amino transferase (ArgD) to convert acetyl glutamate semialdehyde to acetyl ornithine, compared to the endogenous activities thereof, thereby enhancing the biosynthetic pathway of ornithine, a putrescine precursor (Sakanyan V et al., Microbiology. 142:1, 99-108, 1996).

In this case, the ArgF, NCg11221, ODC, ArgC, ArgJ, ArgB and ArgD may have, but are not specifically limited to, the amino acid sequences represented by SEQ ID. NO: 20, 21, 22, 23, 24, 25, 26, respectively, or the amino acid sequences with 80% or more, specifically 90% or more, more specifically 95% or more, and most specifically 97% or more homology with the same.

As used herein, the term "ornithine decarboxylase (ODC)" refers to an enzyme that produces putrescine using ornithine, and the ODC requires pyridoxal phosphate (Pyridoxal 5'-phosphate, PLP) as a coenzyme. The ODC is found in most Gram-negative bacteria and may be found in some of the intestinal bacteria such as *Lactobacillus* of Gram-positive bacteria. *E. coli* has two types of genes encoding ODC, one of which, speC, is expressed continuously at the certain concentration and the other, speF, is expressed under specific conditions (the presence of ornithine at higher than certain concentrations and low pH). Depending on species, some species, like *E. coli*, have two kinds of ODC, and others have only one type. The species such as *Escherichia* sp., *Shigella* sp., *Citrobacter* sp., *Salmonella* sp., and *Enterobacter* sp. have two kinds of ODC (speC, speF), and the strains of *Yersinia* sp., *Klebsiella* sp., *Erwinia* sp., have one kind of ODC (speC). In case of *Lactobacillus*, ODC is expressed in one type of gene (speF), and is known to be induced to be expressed under the conditions of low pH or abundant ornithine and histidine.

ODC activity may be introduced to the recombinant microorganism of genus *Corynebacterium* of the present invention using genes encoding ODC derived from the various species. The polynucleotide encoding the ODC may include, but is not limited to, the polynucleotide encoding the protein consisting of the amino acid sequence represented by SEQ ID NO: 22

and the amino acid sequence of 70% or more, specifically 80% or more, more specifically 90% or more homology with the same.

In addition, the introduction of ornithine decarboxylase (ODC) activity to the microorganisms may be performed by the various methods well known in the art; for example, the method to insert the polynucleotide including a nucleotide sequence encoding ODC to chromosome, the method to introduce the polynucleotide to the microorganisms by introducing to the vector system, the method to insert the promoter which is modified or has improved activity to the upper region of nucleotide sequence encoding ODC, and the method to insert mutation to the nucleotide sequence encoding ODC. More specifically, if the nucleotide sequence encoding ODC is introduced, known CJ7 promoter may be used as a promoter to control the expression of the same.

In addition, the enhancement of the activity of ArgC, ArgJ, ArgB and ArgD can be achieved by 1) an increase of the copy number of polynucleotide encoding the enzyme, 2) a modification of the expression regulatory sequence to increase the polynucleotide expression, 3) a modification of the polynucleotide sequence encoding the enzyme on chromosome to enhance the activity of the enzyme or 4) a combination thereof.

In method 1), the increase of the copy number of polynucleotide encoding the enzyme can be achieved by operably linking the polynucleotide to the vector or by inserting the same to the chromosome of the host cell. More specifically, the copy number of polynucleotide of the host cell can be increased by introducing a vector that is capable of replicating and functioning independently, wherein the polynucleotide encoding the enzyme of the present invention is operably linked, or by introducing the vector capable of inserting the polynucleotide into the chromosome of the host cell, wherein the polynucleotide is operably linked.

As used herein, the term "vector" refers to the DNA construct comprising the nucleotide sequence of the polynucleotide encoding the target protein operably linked to the proper regulatory sequence to express the target protein in the proper host. The regulatory sequence includes the promoter which can initiate transcription, any operator sequence to control the transcription, the sequence to encode the appropriate mRNA ribosome binding site, and the sequence to control the termination of transcription and translation. The vector may be transfected into a suitable host, and then may be replicated or function independently from the host genome, and may be integrated into the genome itself.

In the present invention, any vector which is known in the art may be used without any specific limitation as long as it can be replicated in the host. Examples of commonly used vectors are plasmid, cosmid, virus and bacteriophage in natural state or recombinant state. For example, pWE15, M13, λMBL3, λMBL4, λIXII, λASHII, λAPII, λt10, λt11, Charon4A, and Charon21A can be used as a phage vector or cosmid vector, and pBR system, pUC system, pBluescriptII system, pGEM system, pTZ system, pCL system and pET system can be used as a plasmid vector. The vector which can be used in the present invention is not particularly limited and the known expression vectors can be used. Specifically, pACYC177, pACYC184, pCL, pECCG117, pUC19, pBR322, pMW118, pCC1BAC vectors can be used. Most specifically, pACYC177, pCL, pCC1BAC vectors can be used.

In addition, the vector which can insert the polynucleotide encoding the target protein into chromosome of a host cell may specifically be, for example, a shuttle vector, pECCG112

(Korean Patent Publication No. 1992-0000933) which is able to replicate by itself both in *E. coli* and *Coryneform* bacteria, but is not limited thereto.

In addition, the polynucleotide encoding the target protein in the chromosome may be replaced by a new polynucleotide by using a vector for chromosomal gene insertion. The insertion of the polynucleotide to the chromosome can be achieved by any method known in the art, for example, by homologous recombination. Since the vector of the present invention may be inserted into the chromosome by inducing a homologous recombination, the selection marker may be additionally included to confirm a successful gene insertion into the chromosome. A selection marker is for screening the cells which are transformed with the vector, in other words, for determining whether the target polynucleotide is inserted. The markers that provide selectable phenotypes such as drug resistance, auxotrophy, resistance to toxic agents or expression of surface proteins may be used. In an environment treated with a selective agent, only the cells expressing the selection marker can survive or cells show a different phenotype, and thus the successfully transformed cells can be selected through this method.

As used herein, the term "transformation" refers to the introduction of the vector comprising a polynucleotide encoding the target protein into the host cell so that the protein can be expressed in the cell. The transformed polynucleotide includes all polynucleotide which encode target proteins that can be expressed in the host cell regardless of the location, whether it is inserted into the chromosome of the host cell or located outside the chromosome. In addition, the polynucleotide includes DNA and RNA encoding the target protein. The polynucleotide may be introduced in any form as long as it can be introduced into the host cell and expressed. For example, the polynucleotide can be introduced into a host cell in a form of an expression cassette which is gene construct, comprising all the required elements for self-expression. The expression cassette typically includes a promoter operably linked to the polynucleotide, transcription termination signal, ribosomal binding site, and translation termination signal. The expression cassette may be the form of expression vector capable of self-replication. In addition, the polynucleotide may be introduced into a host cell in its own form and operably linked to the sequences required for the expression of host cell.

As used herein, the term "operably linked" refers to the functional connection between the promoter sequence initiating or mediating the transcription of polynucleotide encoding the target protein and the polynucleotide.

In addition, the method 2) modification of the expression regulatory sequence to increase the expression of the polynucleotide in the present invention may be performed by inducing the mutation of the sequence through deletion, insertion, conservative or non-conservative substitution of nucleotide sequence or a combination thereof, or by substitution by the nucleotide sequence with enhanced activity. The expression regulatory sequence includes promoter, operator sequence, sequence encoding ribosomal binding sites, and sequence to control the termination of transcription and translation.

A strong heterologous promoter may be linked to the upper of expression unit of the polynucleotide instead of original promoters. An example of a strong promoter is pcj7 promoter, lysCP1 promoter, EF-Tu promoter, groEL promoter, aceA or aceB promoter, etc., and more specifically lysCP1 promoter or pcj7 promoter derived from *Corynebacterium* is operably linked to enhance the expression of polynucleotide encoding the enzyme. Herein, lysCP1 promoter, which is an improved

promoter through substitution of the nucleotide sequence of the promoter region of polynucleotide encoding aspartate kinase and aspartate semialdehyde dehydrogenase, is strong enough to increase the activity of the corresponding enzyme by 5 times compared to the wild type through enhancement of expression of aspartate kinase gene (International Patent Publication No. 2009-096689). In addition, the pcj7 promoter was identified to be expressed in *Corynebacterium ammoniagenes* and *Escherichia* and to have a strong promoter activity, and can be expressed in *Corynebacterium glutamicum* as well in high intensity (Korean Patent No. 0620092).

In addition, the method 3) modification of the polynucleotide sequence on chromosome may be performed, but are not specifically limited to, by inducing the mutation of the sequence through deletion, insertion, conservative or non-conservative substitution of nucleotide sequence or a combination thereof to enhance the activity of the sequence, or by substitution by the nucleotide sequence having enhanced activity.

The microorganism in the present invention, which is a microorganism having the ability to produce putrescine, includes prokaryotic microorganism, wherein the protein comprising amino acid sequence represented by in SEQ ID NO: 17 or SEQ ID NO: 19 is expressed, and may be, for example, the microorganism of *Escherichia* sp., *Shigella* sp., *Citrobacter* sp., *Salmonella* sp., *Enterobacter* sp., *Yersinia* sp., *Klebsiella* sp., *Erwinia* sp., *Corynebacterium* sp., *Brevibacterium* sp., *Lactobacillus* sp., *Sphenomanas* sp., and *Vibrio* sp.

The microorganism in the present invention is specifically the microorganism of genus *Corynebacterium* and may more specifically be of *Corynebacterium glutamicum*.

An embodiment of the present invention, the microorganism of genus *Corynebacterium* of accession number KCCM11138P (Korean Patent laid-open No. 2012-0064046), which has the ability to produce putrescine in a high concentration through enhanced putrescine-biosynthesis pathway, was modified. Specifically, the putrescine-producing strain KCCM11138P is the putrescine-overproducing strain, wherein the gene encoding ornithine carbamoyltransferase (ArgF) for accumulating ornithine and the gene encoding glutamate exporter (NCg11221; SEQ ID NO: 21) for increasing intracellular glutamate are deleted from ATCC13032 strains, the gene encoding ornithine decarboxylase (spec) is introduced, and the expression level of ornithine biosynthesis genes (argCJBD) is increased.

Another embodiment of the present invention, *Corynebacterium glutamicum* ATCC13869-based putrescine-producing strain DAB12-a was modified. The strain ATCC13869 was based on the same genotype as the KCCM11138P, which is putrescine-producing strain, based on *Corynebacterium glutamicum* ATCC13032. Specifically, putrescine-producing strain DAB12-a is from ATCC13869 strain obtained from American Type Culture Collection (ATCC), wherein the gene encoding ornithine carbamoyltransferase (ArgF) and the gene encoding the protein NCg11221 (SEQ ID NO: 21) to export glutamate are deleted, the gene (spec) encoding ornithine decarboxylase (ODC) derived from *E. coli* is introduced, and the promoter of ornithine biosynthesis gene operon (argCJBD) is replaced with the improved promoter.

According to one embodiment of the present invention, a microorganism of genus *Corynebacterium* (KCCM11138P) has an ability to produce putrescine, which is prepared by deletion of the gene encoding ornithine carbamoyl transferase (ArgF) and the gene encoding the glutamate exporter (NCg11221; SEQ ID NO: 21) involved in glutamate export, replacement of the own promoter of ArgCJBD gene cluster

encoding an enzyme involved in the synthesis of ornithine from glutamate, and introduction of the gene (spec) encoding ornithine decarboxylase (ODC) into the chromosome in the wild-type *Corynebacterium glutamicum* ATCC13032. Based on KCCM11138P, a clone (A15) growing well in a medium containing high concentration of putrescine was selected, and it was confirmed that the selected A15 includes genes encoding NCg10100 (SEQ ID NO: 27), NCg10101 (SEQ ID NOS: 17 or 19), NCg10102 (SEQ ID NO: 29), NCg10103 (SEQ ID NO: 30) and NCg10104 (SEQ ID NO: 31) (Example 1). In addition, the microorganism grows in the medium containing high concentration of putrescine due to the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) among the five types of genes (Example 2). As regards character of the gene encoding NCg10101 (SEQ ID NOS: 17 or 19), it was confirmed that putrescine production was reduced in a strain in which the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) is over-expressed (Example 3), and putrescine production was increased in a strain in which the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) is deleted (Example 4).

Accordingly, the present inventors named the *Corynebacterium glutamicum* strain having an enhanced ability to produce putrescine, which is prepared by removing the NCg10101 (SEQ ID NOS: 17 or 19) gene in the putrescine-producing strain KCCM 11138P, as *Corynebacterium glutamicum* CC01-0244, and deposited in the Korean Culture Center of Microorganisms (hereinafter, abbreviated to "KCCM") on Dec. 26, 2011, with Accession No. KCCM11241P.

In another aspect of the present invention to achieve the above objectives, the present invention relates to a method for producing putrescine, comprising the steps of:

culturing the microorganism of genus *Corynebacterium* having an enhanced ability to produce putrescine, which is modified to have the weakened activity of NCg10101 protein (SEQ ID NOS: 17 or 19) having an amino acid sequence represented by SEQ ID NO. 17 or SEQ ID NO. 19; and isolating putrescine from the culture broth obtained in the above step.

The culturing process in the present invention may be carried out in appropriate medium and under culturing conditions known in the art. Those skilled in the art can easily adjust and use the culturing process depending on selected strains. An example of the culturing process includes batch, continuous and fed-batch type cultures, but is not limited thereto. The culture medium may have to appropriately satisfy the requirements of a specific strain.

The culture medium may have to appropriately satisfy the requirements of specific strains. Culture media for various microorganisms are disclosed (for example, "Manual of Methods for General Bacteriology" from American Society for Bacteriology (Washington D.C., USA, 1981)). As a source of carbon in the medium, sugar and carbohydrates (e.g., glucose, sucrose, lactose, fructose, maltose, molasses, starch, and cellulose), butterfat and fat (e.g., soybean oil, sunflower seed oil, peanut oil and coconut oil), fatty acid (e.g., palmitic acid, stearic acid and linoleic acid), alcohol (e.g., glycerol and ethanol) and organic acid (e.g., acetic acid), etc. may be used. These substances may be used individually or as a mixture. As a source of nitrogen, nitrogen-containing organic compound (e.g., peptone, yeast extract, beef extract, malt extract, corn steep liquor, soybean meal powder and urea) or inorganic compound (e.g., ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate and ammonium nitrate) may be used and these substances also may be used individually or as a mixture. As a source of phosphorus, potassium dihydrogen phosphate or

dipotassium hydrogen phosphate or the corresponding sodium-containing salt may be used. In addition, the culture medium may comprise metal salt (e.g., magnesium sulfate or iron sulfate) which is essential for the growth, and finally, essential growth-promoting substances such as amino acids and vitamins, may be used in addition to the above-mentioned substances. The appropriate precursor may be added in addition to the culture medium. The feed substance may be provided in the culture at once or adequately while culturing.

The pH of the culture may be adjusted by a proper basic compound (e.g., sodium hydroxide, potassium hydroxide or ammonia) or acidic compound (e.g., phosphoric acid or sulfuric acid). Foaming may be adjusted by an anti-foaming agent such as fatty acid polyglycoester. Aerobic condition of the culture may be maintained by introducing oxygen or oxygen-containing gas mixtures, for example, air. Culturing temperature may be typically 20 to 45° C., specifically 25 to 40° C. Culturing may be continued until the production of putrescine reaches the desired maximum, it may be usually achieved in 10 to 160 hours. Putrescine may be released into culture medium, or contained in the cell.

For the method for collecting and recovering the produced putrescine in the culturing process of the present invention, the target substance may be recovered from the culture medium using the appropriate known method in the art depending on the culture method, for example, batch, continuous or fed-batch type culture.

Mode for Invention

Hereinafter, the present invention will be described in more detail with the following Examples. However, these Examples are for illustrative purposes only, and the invention is not intended to be limited by these Examples.

Example 1

Library Preparation for Selection of Effective Genes for Putrescine Biosynthesis and Selection of Clones

In order to screen effective genes for putrescine biosynthesis from the chromosome of the wild-type *Corynebacterium* strain, a chromosome library of the wild-type *Corynebacterium* strain was prepared. In detail, the chromosome extracted from the wild-type *Corynebacterium glutamicum* ATCC 13032 strain was randomly cleaved with the restriction enzyme Sau3AI, and fragments of 5 to 8 kb were selected therefrom, and then cloned into an *E. coli*-*Corynebacterium* shuttle vector pECCG122 (Korean Patent laid-open No. 1992-0000933) to prepare a chromosome library.

In order to select effective genes for putrescine biosynthesis from the *Corynebacterium* chromosome library thus prepared, colonies growing in a medium containing high concentration of putrescine were obtained.

Meanwhile, the libraries were introduced into a microorganism of *Corynebacterium* genus (KCCM11138P) having an ability to produce putrescine, so as to prepare each of transformants. The transformants which were able to grow in a minimal medium containing 0.35 M putrescine (10 g/l of glucose, 0.4 g/l of MgSO₄·7H₂O, 4 g/l of NH₄Cl, 1 g/l of KH₂PO₄, 1 g/l of K₂HPO₄, 2 g/l of urea, 10 mg/l of FeSO₄·7H₂O, 1 mg/l of MnSO₄·5H₂O, 5 mg/l of nicotinamide, 5 mg/l of thiamine hydrochloride, 0.1 mg/l of biotin, 1 mM arginine, 25 mg/l of kanamycin, 0.35 M putrescine, pH 7.0) were selected. The strain KCCM11138P is disclosed in a patent applied by the present inventors (Korean Patent laid-open No. 2012-0064046), which was prepared by deleting

genes encoding ornithine carbamoyltransferase (argF) and glutamate exporter (NCg11221; SEQ ID NO: 21) in the chromosome of the wild type *Corynebacterium glutamicum* strain ATCC 13032, introducing a gene (spec) encoding ornithine decarboxylase (ODC) derived from the wild type *E. coli* W3110 strain into the chromosome, and replacing the promoter of argCJBD gene cluster encoding the enzyme involved in the synthesis of ornithine from glutamate, so as to prepare each of transformants. As a result, 275 colonies were selected, and colonies growing well in the medium containing high concentration of putrescine were secondarily identified. Each library clone was obtained and introduced into the putrescine strain again. Thereafter, colonies growing well in the medium containing high concentration of putrescine were identified and thus a clone (A15) was finally selected. This selected clone was identified by sequencing. As a result, it was confirmed that the clone comprises total 5 ORFs that encode NCg10100 (SEQ ID NO: 27), NCg10101 (SEQ ID NOS: 17 or 19), NCg10102 (SEQ ID NO: 29), NCg10103 (SEQ ID NO: 30) and NCg10104 (SEQ ID NO: 31), of which 436 amino acids at the N-terminus were removed (FIG. 1). FIG. 1 is a schematic diagram showing the relative positions of genes encoding NCg10100 (SEQ ID NO: 27), NCg10101 (SEQ ID NOS: 17 or 19), NCg10102 (SEQ ID NO: 29), NCg10103 (SEQ ID NO: 30) and NCg10104 (SEQ ID NO: 31), which are on the chromosome of the wild type *Corynebacterium glutamicum* ATCC 13032 strain.

Example 2

Identification of Effective Genes for Putrescine Synthesis in A15 Clone

Example 2-1

Cloning of 5 Genes in A15 Clone and Preparation of a Transformant

The nucleotide sequence of the A15 clone obtained in Example 1 was already known. Based on the nucleotide sequence of ATCC13032 strain previously reported, NCg10100-F and NCg10100-R represented by SEQ ID NOS. 1 and 2 as primers for amplification of the gene that encodes NCg10100 (SEQ ID NO: 27), NCg10100-R and tNCg10100-F represented by SEQ ID NOS. 2 and 3 as primers for amplification of the gene that encodes tNCg10100 (SEQ ID NO: 28) of which 436 amino acids at the N-terminus were removed, NCg10101-F and NCg10101-R represented by SEQ ID NOS. 4 and 5 as primers for amplification of gene that encodes NCg10101 (SEQ ID NOS: 17 and 19), NCg10102-F and NCg10103-R represented by SEQ ID NOS. 6 and 7 as primers for amplification of both the genes that encode NCg10102 (SEQ ID NO: 29) and NCg10103 (SEQ ID NO: 30), and NCg10104-F and NCg10104-R represented by SEQ ID NOS. 8 and 9 as primers for amplification of the gene that encodes NCg10104 (SEQ ID NO: 31) were constructed. In addition, P(CJ7)-F and P(CJ7)-R represented by SEQ ID NOS. 10 and 11 as primers for amplification of the expression promoter P(CJ7) (or pcj7) (Korean Patent No. 10-0620092) were constructed (Table 1).

Thereafter, PCR was carried out using the chromosome of ATCC 13032 strain as a template and each of the primer represented by SEQ ID NOS. 1 to 9 (denaturation at 95° C. for 30 seconds, annealing at 50° C. for 30 seconds, and extension at 72° C. for 1 minute~1 minute 30 seconds, 25 cycles), so as to amplify 5 types of gene fragments. In addition, PCR was carried out using the chromosome of *Corynebacterium*

11

ammoniagenes as a template and primers represented by SEQ ID NOS. 10 and 11 so as to amplify the promoter fragment.

5 genes cleaved with KpnI and XbaI, and CJ7 promoter cleaved with EcoRV and KpnI were ligated into an expression vector pHC139T (Korean Patent No. 10-0860932) cleaved with EcoRV and XbaI, so as to prepare total 5 types of expression vectors, pHC139T-P(CJ7)-NCg10100, pHC139T-P(CJ7)-tNCg10100, pHC139T-P(CJ7)-NCg10101, pHC139T-P(CJ7)-NCg10102-NCg10103, and pHC139T-P(CJ7)-NCg10104.

TABLE 1

Primers for preparation of strains expressing 5 genes contained in A15 clone	
NCg10100-F (SEQ ID NO. 1)	GCGCAT ATGAGCTCAAC AACCTCAAAACC
NCg10100-R (SEQ ID NO. 2)	GCGTCTAGA TTATCCTT CGAGGAAGATCGCAG
tNCg10100-F (SEQ ID NO. 3)	GCGCAT ATGTGGACGCT GATGGCTGC
NCg10101-F (SEQ ID NO. 4)	GCGCAT ATGAGTACTGA CAATTTTCTCCAC
NCg10101-R (SEQ ID NO. 5)	GCGTCTAGA CTAAGCCA AATAGTCCCCTAC
NCg10102-F (SEQ ID NO. 6)	GCGCAT ATGGATGAACG AAGCCGTTTG
NCg10103-R (SEQ ID NO. 7)	GCGTCTAGATTAATCAAT GAAGACGAATAAATTC
NCg10104-F (SEQ ID NO. 8)	GCGCATATGGCGGGTGAC AAATTGTGG
NCg10104-R (SEQ ID NO. 9)	GCGTCTAGATTAGGACAG TTCCGCTGGAGC
P(CJ7)-F (SEQ ID NO. 10)	CAGATATCGCCGGCATAG CCTACCGATG
P(CJ7)-R (SEQ ID NO. 11)	GCGTCTAGAGATATCAGT GTTTCCTTCG

5 types of the expression vectors thus prepared and a control group pHC139T were introduced into the KCCM11138P strain of Example 1 by electroporation, and then spread on BHIS plates containing 25 µg/ml kanamycin to select transformants.

Example 2-2

Search of Effective Genes for Putrescine

From the total 6 types of the transformants obtained in Example 2-1, transformants growing well in the medium containing high concentration of putrescine were selected in the same manner as in Example 1 (FIG. 2). FIG. 2 is the test result of comparing growth between the transformants prepared in the present invention, in which 1, 2, 3, 4, 5 and 6 represent strains introduced with the 6 types of expression vectors, pHC139T, pHC139T-P(CJ7)-NCg10100, pHC139T-P(CJ7)-tNCg10100, pHC139T-P(CJ7)-NCg10101, pHC139T-P(CJ7)-NCg10102-NCg10103 and pHC139T-P(CJ7)-NCg10104, respectively. As shown in FIG. 2, only the transformant (No. 4) introduced with pHC139T-P(CJ7)-NCg10101 showed excellent growth in the medium contain-

12

ing high concentration of putrescine, and thus NCg10101 (SEQ ID NOS: 17 or 19) was selected as the effective gene for putrescine biosynthesis.

Example 3

Evaluation of the Ability to Produce Putrescine in NCg10101-Overexpressing Strain

The ability to produce Putrescine of the strain overexpressing the NCg10101 (SEQ ID NOS: 17 or 19) gene which was identified as the effective gene in Example 2 was evaluated. A strain for evaluation was prepared by introducing pHC139T-P(CJ7)-NCg10101 into the putrescine-producing strain KCCM11138P.

pHC139T-P(CJ7)-NCg10101 prepared in Example 2-1 and pHC139T vector as a control group were introduced into the putrescine-producing strain KCCM 11138P by electroporation, and then spread on BHIS plates containing 25 µg/ml kanamycin to select transformants. The transformants were named as KCCM 11138P/pHC139T, and KCCM 11138P/pHC139T-P(CJ7)-NCg10101, respectively. These two transformants thus selected were cultured in CM plates containing 1 mM arginine (1% glucose, 1% polypeptone, 0.5% yeast extract, 0.5% beef extract, 0.25% NaCl, 0.2% urea, 100 µl of 50% NaOH, 2% agar, pH 6.8 per 1 L) at 30° C. for 24 hours, and then a loop of cell culture was inoculated in 25 ml of titer medium of Table 2 containing 25 µg/ml kanamycin, and cultured with shaking at 200 rpm at 30° C. for 96 hours. All of the prepared strains were cultured with addition of 1 mM arginine in the medium during fermentation.

TABLE 2

Composition	Concentration (per 1 L)
Glucose	8%
Soybean protein	0.25%
Corn steep solids	0.5%
(NH ₄) ₂ SO ₄	4%
Urea	0.15%
KH ₂ PO ₄	0.1%
MgSO ₄ ·7H ₂ O	0.05%
Biotin	100 µg
Thiamine Hydrochloride	3000 µg
Calcium-Panthotenic Acid	3000 µg
Nicotinamide	3000 µg
CaCO ₃	5%

As a result, as shown in Table 3, when NCg10101 (SEQ ID NOS: 17 or 19) was overexpressed, putrescine production was reduced.

TABLE 3

Strain type	Putrescine (g/L)
KCCM 11138P/pHC139T	9.5
KCCM 11138P/pHC139T-P(CJ7)-NCg10101	5.1

13

Example 4

Evaluation of the Ability to Produce Putrescine in
NCg10101-Deleted Strain

Example 4-1

Preparation of NCg10101-Deleted Strain in ATCC
13032-Based Putrescine-Producing Strain

NCg10101 (SEQ ID NOS: 17 or 19) overexpression increased cell growth in the medium containing high concentration of putrescine, but decreased putrescine production according to Example 3. On the basis of this result, the effect of NCg10101 (SEQ ID NOS: 17 or 19) deletion on the ability to produce putrescine was examined.

In detail, based on the NCg10101 nucleotide sequence of ATCC 13032 strain (SEQ ID NOS: 16 or 18), NCg10101-del-F1_BamHI and NCg10101-del-R1_Sall represented by SEQ ID NOS. 12 and 13 as primers were constructed to obtain a homologous recombinant fragment of the N-terminal region of NCg10101 (SEQ ID NOS: 17 or 19). NCg10101-del-F2_Sall and NCg10101-del-R2_XbaI represented by SEQ ID NOS. 14 and 15 as primers were constructed to obtain a homologous recombinant fragment of the C-terminal region of NCg10101 (SEQ ID NOS: 17 or 19) (Table 4). The fragments of the N-terminal and C-terminal regions of NCg10101 gene (SEQ ID NOS: 16 or 18) were prepared by PCR using the two pairs of the primers. The PCR products were treated with BamHI & Sall and Sall & XbaI, respectively and cloned into a pDZ vector treated with BamHI & XbaI. The cloned plasmid was named as pDZ-NCg10101(K/O).

TABLE 4

Primers for preparation of NCg10101-deleted strains	
NCg10101-del-F1_BamHI (SEQ ID NO. 12)	CGGGATCC CGGATTCCTGCGATCATTG
NCg10101-del-R1_Sall (SEQ ID NO. 13)	ACGCGTCGAC CAGTCGACGGAACCTGTGGAG
NCg10101-del-F2_Sall (SEQ ID NO. 14)	ACGCGTCGAC GGCAACGACTCCGAAACCTTC
NCg10101-del-R2_XbaI (SEQ ID NO. 15)	CTAGTCTAGA CTGGATCCTCATGAATGCGC

The pDZ-NCg10101(K/O) vector prepared for obtaining the KCCM 11138P ΔNCg10101 strain was introduced into KCCM 11138P strain by electroporation, and then spread on the BHIS plate containing 25 μg/ml kanamycin. The successful insertion of the vector in the chromosome was confirmed by observing whether the colony was blue on the solid medium containing X-gal (5-bromo-4-chloro-3-indolyl-β-D-galactoside). The primary chromosome inserted strain was shaking-cultured in a nutrient medium (30° C., 8 hours), was then diluted from 10⁻⁴ to 10⁻¹⁰, and spread on the solid medium containing X-gal. While a majority of colonies appeared as blue colony, a low proportion of colonies appeared as white colonies. The NCg10101 (SEQ ID NOS: 17 or 19) gene-deleted strains were finally selected by double crossover with the white colonies, and identified by PCR using the primers represented by SEQ ID NOS. 12 and 15. The variant thus identified was named as KCCM 11138P ΔNCg10101.

14

Example 4-2

Preparation of NCg10101-Deleted Strain in ATCC
13869-Based Putrescine-Producing Strain

Corynebacterium glutamicum ATCC13869-based putrescine-producing strain DAB12-a (argF-deleted, NCg11221 (SEQ ID NOS: or 19)-deleted, *E. coli* speC-introduced, and arg operon-argCJBD promoter-substituted strain), which has the same genotype as that of the putrescine-producing strain KCCM11138P based on *Corynebacterium glutamicum* ATCC13032, was used to prepare NCg10101 (SEQ ID NOS: 17 or 19)-deleted strains.

In detail, in order to identify the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) derived from *Corynebacterium glutamicum* ATCC13869 and the amino acid sequence of the protein expressed therefrom, PCR was carried out using the genomic DNA of *Corynebacterium glutamicum* ATCC13869 as a template and a pair of primers, SEQ ID NOS. 12 and 15 (NCg10101-del-F1_BamHI, NCg10101-del-R2_XbaI). Here, PCR reaction was carried out with 30 cycles of denaturation at 95° C. for 30 seconds, annealing at 53° C. for 30 seconds, and extension at 72° C. for 2 minutes and 30 seconds. The PCR products were separated by electrophoresis and their sequences were analyzed. Through sequence analysis, it was identified that the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) derived from *Corynebacterium glutamicum* ATCC13869 includes a nucleotide sequence represented by SEQ ID NO. 18 and the protein encoded thereby includes an amino acid sequence represented by SEQ ID NO. 19. When the amino acid sequences of NCg10101 (SEQ ID NOS: 17 or 19) derived from *Corynebacterium glutamicum* ATCC13032 and that of NCg10101 (SEQ ID NOS: 17 or 19) derived from *Corynebacterium glutamicum* ATCC13869 were compared, they showed 98% sequence homology.

In order to delete the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) derived from *Corynebacterium glutamicum* ATCC13869, the region of N-terminal and C-terminal of NCg10101 gene (SEQ ID NOS: 16 or 18) were amplified by PCR using a genomic DNA of *Corynebacterium glutamicum* ATCC13869 as a template and two pairs of primers listed in Table 4 in the same manner as Example <4-1>. Then, the PCR products were treated with BamHI & Sall and Sall & XbaI, respectively and then cloned into the pDZ vector treated with BamHI & XbaI, thereby constructing a plasmid pDZ-2'NCg10101(K/O).

The plasmid pDZ-2'NCg10101(K/O) was transformed into *Corynebacterium glutamicum* DAB12-a in the same manner as in Example <4-1>, and the strain in which the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) is deleted was selected. The selected *Corynebacterium glutamicum* variant was named as DAB12-a ΔNCg10101.

Example 4-3

Evaluation of the Ability to Produce Putrescine in
NCg10101-Deleted Strain

In order to investigate the effect of NCg10101 (SEQ ID NOS: 17 or 19) deletion on the ability to produce putrescine in the putrescine-producing strain, the *Corynebacterium glutamicum* variants prepared in Examples <4-1> and <4-2> was compared.

In detail, the ability to putrescine in two types of *Corynebacterium glutamicum* variants (KCCM11138P ΔNCg10101 and DAB12-a ΔNCg10101) was evaluated in the same manner as in example 3. As shown in the following Table 5,

15

putrescine production was found to be increased by NCg10101 (SEQ ID NOS: 17 or 19) deletion.

TABLE 5

Strain type	Putrescine (g/L)
KCCM 11138P	9.8
KCCM 11138P ΔNCg10101	11.3
DAB12-a	10.1
DAB12-a ΔNCg10101	11.0

Taken together, the results of Examples 3 and 4 show that putrescine production was decreased by overexpression of the gene encoding NCg10101 (SEQ ID NOS: 17 or 19) and increased by deletion of the gene in the wild type *Corynebacterium glutamicum* strain, indicating that NCg10101 (SEQ ID NOS: 17 or 19) directly affects putrescine biosynthesis.

Accordingly, the present inventors named the *Corynebacterium glutamicum* strain having an improved ability to pro-

16

duce putrescine, which was prepared by deleting the NCg10101 gene (SEQ ID NOS: 16 or 18) in the putrescine-producing strain KCCM 11138P in the above Example, as *Corynebacterium glutamicum* CC01-0244, and deposited in Korean Culture Center of Microorganisms (hereinafter, abbreviated to as “KCCM”) which is international depositary authority under the Budapest Treaty on Dec. 26, 2011, with Accession No. KCCM11241P.

Based on the above descriptions, those skilled in the art will understand that the present invention may be conducted in other forms without changing the technical idea or essential technical features. In this regard, the Examples described above are to illustrate the invention in all respects, but not to limit the scope of the invention. It shall be understood that the scope of the present invention comprises any changes or modified forms derived from the meaning, scope and equivalent concept of the following claims rather than the detailed descriptions in the above.

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Asn Ala Ser Thr Lys Gly Glu Ser Pro Asp Tyr Pro Gly Gln Gln Val
35        40        45
Ile Trp Arg Leu Ile Gln Glu Ala Gly Glu Ser Leu Arg Asp Glu Leu
50        55        60
Arg Thr Leu Ala Phe Thr Leu His Asp His Pro Glu Glu Ala Phe Glu
65        70        75        80
Glu Val Phe Ala Thr Glu Glu Ile Thr Lys Leu Leu Gln Asn His Gly
85        90        95
Phe Glu Val Gln Ser Gly Val Tyr Gly Val Lys Thr Ala Leu Glu Thr
100       105       110
Ser Phe Glu Thr Pro Gly Tyr Asp Pro Ala Gln His Pro Ser Ile Ala
115       120       125
Ile Leu Ala Glu Tyr Asp Ala Leu Pro Glu Ile Gly His Ala Cys Gly
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His Asn Ile Ile Ala Ala Ala Gly Val Gly Ala Phe Leu Ala Val Thr
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Asn Met Ile Lys Thr Ala Glu Val Lys Gly Val Asp His Leu Asp Phe
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Glu Gly Arg Ile Val Leu Leu Gly Thr Pro Ala Glu Glu Gly His Ser
180       185       190
Gly Lys Glu Tyr Met Ile Arg Asn Gly Ala Phe Asp Gly Ile Asp Ala
195       200       205
Ser Ile Met Met His Pro Phe Gly Phe Asp Leu Ala Glu His Val Trp
210       215       220
Val Gly Arg Arg Thr Met Thr Ala Thr Phe His Gly Val Ser Ala His
225       230       235       240
Ala Ser Ser Gln Pro Phe Met Gly Lys Asn Ala Leu Asp Ala Ala Ser

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245								250				255			
Leu	Ala	Tyr	Gln	Gly	Phe	Gly	Val	Leu	Arg	Gln	Gln	Met	Pro	Pro	Ser
			260				265						270		
Asp	Arg	Leu	His	Ala	Ile	Ile	Thr	Glu	Gly	Gly	Asn	Arg	Pro	Ser	Ile
			275				280			285					
Ile	Pro	Asp	Thr	Ala	Thr	Met	Ser	Leu	Tyr	Val	Arg	Ser	Leu	Leu	Pro
			290	295						300					
Glu	Ala	Leu	Lys	Asp	Ile	Ser	Lys	Arg	Val	Asp	Asp	Val	Leu	Asp	Gly
305			310						315			320			
Ala	Ala	Leu	Met	Ala	Gly	Val	Gly	Val	Glu	Lys	Gln	Trp	Asp	Val	His
			325						330			335			
Pro	Ala	Ser	Leu	Pro	Val	Arg	Asn	Asn	His	Val	Leu	Ala	Arg	Arg	Trp
			340			345						350			
Ala	Lys	Thr	Gln	Asn	Leu	Arg	Gly	Arg	Thr	Ala	Leu	Ser	Glu	Gly	Ile
			355			360						365			
Leu	Pro	Asp	Thr	Leu	Ala	Ala	Ser	Thr	Asp	Phe	Gly	Asn	Val	Ser	His
370			375						380						
Leu	Val	Pro	Gly	Ile	His	Pro	Met	Val	Lys	Ile	Ser	Pro	Glu	Asn	Val
385			390						395			400			
Ala	Leu	His	Thr	Lys	Glu	Phe	Ala	Ala	Tyr	Ala	Arg	Thr	Glu	Glu	Ala
			405						410			415			
Ile	Asp	Ala	Ala	Val	Asp	Ala	Ala	Ile	Gly	Leu	Ala	Gln	Val	Ala	Val
			420			425						430			
Asp	Ala	Leu	Ala	Asp	Pro	Gln	Met	Leu	Ile	Asp	Ala	Thr	Leu	Glu	Phe
			435			440						445			
Thr	Asn	Ser	Gly	Asp	Val	Leu	Lys	Val	Gly	Asp	Tyr	Leu	Ala		
450			455						460						

<210> SEQ ID NO 18

<211> LENGTH: 1389

<212> TYPE: DNA

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 18

```

atgagtactg acaatttttc tccacaagtt ccgctcgactg tgtatttgga ttacatggag      60
caagggattg tcgcgcgtaa agcggaggca gaatctaacg ccagcacgca gggggagagc      120
ccggattatc caggccagca ggttatttgg cgcctgatcc aggaagcagg ggagtcgttg      180
cgtgatgaac tgcgcacact ggttttcacg ctgcacgacc atccggaaga agcgttcgag      240
gaggtgttcg ccaccgagga aatcacaaaa cttctgcaaa atcatggttt tgaggttcag      300
agtggagttt atggtgttaa aaccgctcta gaaactagtt ttgaaacccc tggttatgat      360
ccagcgcagc acccaagcat tgcgatcttg gcggaatacg atgcccttcc agagatcggc      420
catgcgtgcg ggcacaatat catcgcagca gctggtgttg gtgcattttt ggctgtcacc      480
aacatgatca aaaatgccga agtgaaaggc gtggatcacc tcgactttga aggccggatc      540
gtgctgttgg gaacacctgc cgaagaaggg cattccggca aggaatacat gatccgaaat      600
ggcgcatctg atggcattga tgcattccatc atgatgcacc cctttggctt cgatctggcg      660
gaacatgttt ggggtggcag gcgcactatg acggcgacgt tccacggtgt ctctgcacac      720
gcgtcttcgc agcctttcat gggtaaaaat gccctcgacg ctgcaagttt ggcgtaccag      780
ggcttcggag ttttgcgtca gcaaatgccca ccgagcagacc gccttcacgc cattattacg      840
gaaggcggaa accggccaag catcattcca gacactgcaa cgatggcggt gtatgtgcgt      900

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tccctgctgc cggaagcact caaagacata tcgaaacgcg tggatgatgt gctcgatggg 960
gcggccttga tggcgggggt tggcgctcgaa aagcaatggg atgtgcaccc agctagcttg 1020
cccggtgcga acaatcatgt gttggcgcg cggtgggcaa aaacgcagaa tctgcgtggt 1080
cgaacggcgc tttcggaggg cattttgcc gacactctgg cagcatcgac tgattttggc 1140
aatgtctcgc acctgattcc gggcattcat ccgatggta aaatttctcc ggaaaacgtt 1200
gcgctccaca ccaaggaatt cgccgcttat gcgcgcacgg aagaggccat cgacgcagcc 1260
gtcgacgcgc caatcgggct ggcgcaagtc gccgttgacg cgcttgacaga tccgcaaatg 1320
cttatcgacg cgaccctcga gtccaccaac tccggcgcca tgcttaaagc gggagactat 1380
ttggttag 1389

```

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<210> SEQ ID NO 19
<211> LENGTH: 462
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum

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<400> SEQUENCE: 19

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```

Met Ser Thr Asp Asn Phe Ser Pro Gln Val Pro Ser Thr Val Tyr Leu
1      5      10      15
Asp Tyr Met Glu Gln Gly Ile Val Ala Arg Lys Ala Glu Ala Glu Ser
20     25     30
Asn Ala Ser Thr Gln Gly Glu Ser Pro Asp Tyr Pro Gly Gln Gln Val
35     40     45
Ile Trp Arg Leu Ile Gln Glu Ala Gly Glu Ser Leu Arg Asp Glu Leu
50     55     60
Arg Thr Leu Ala Phe Thr Leu His Asp His Pro Glu Glu Ala Phe Glu
65     70     75     80
Glu Val Phe Ala Thr Glu Glu Ile Thr Lys Leu Leu Gln Asn His Gly
85     90     95
Phe Glu Val Gln Ser Gly Val Tyr Gly Val Lys Thr Ala Leu Glu Thr
100    105    110
Ser Phe Glu Thr Pro Gly Tyr Asp Pro Ala Gln His Pro Ser Ile Ala
115    120    125
Ile Leu Ala Glu Tyr Asp Ala Leu Pro Glu Ile Gly His Ala Cys Gly
130    135    140
His Asn Ile Ile Ala Ala Ala Gly Val Gly Ala Phe Leu Ala Val Thr
145    150    155    160
Asn Met Ile Lys Asn Ala Glu Val Lys Gly Val Asp His Leu Asp Phe
165    170    175
Glu Gly Arg Ile Val Leu Leu Gly Thr Pro Ala Glu Glu Gly His Ser
180    185    190
Gly Lys Glu Tyr Met Ile Arg Asn Gly Ala Phe Asp Gly Ile Asp Ala
195    200    205
Ser Ile Met Met His Pro Phe Gly Phe Asp Leu Ala Glu His Val Trp
210    215    220
Val Gly Arg Arg Thr Met Thr Ala Thr Phe His Gly Val Ser Ala His
225    230    235    240
Ala Ser Ser Gln Pro Phe Met Gly Lys Asn Ala Leu Asp Ala Ala Ser
245    250    255
Leu Ala Tyr Gln Gly Phe Gly Val Leu Arg Gln Gln Met Pro Pro Ser
260    265    270
Asp Arg Leu His Ala Ile Ile Thr Glu Gly Gly Asn Arg Pro Ser Ile
275    280    285

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Ile Pro Asp Thr Ala Thr Met Ala Leu Tyr Val Arg Ser Leu Leu Pro
 290                295                300

Glu Ala Leu Lys Asp Ile Ser Lys Arg Val Asp Asp Val Leu Asp Gly
 305                310                315                320

Ala Ala Leu Met Ala Gly Val Gly Val Glu Lys Gln Trp Asp Val His
                325                330                335

Pro Ala Ser Leu Pro Val Arg Asn Asn His Val Leu Ala Arg Arg Trp
                340                345                350

Ala Lys Thr Gln Asn Leu Arg Gly Arg Thr Ala Leu Ser Glu Gly Ile
                355                360                365

Leu Pro Asp Thr Leu Ala Ala Ser Thr Asp Phe Gly Asn Val Ser His
 370                375                380

Leu Ile Pro Gly Ile His Pro Met Val Lys Ile Ser Pro Glu Asn Val
 385                390                395                400

Ala Leu His Thr Lys Glu Phe Ala Ala Tyr Ala Arg Thr Glu Glu Ala
                405                410                415

Ile Asp Ala Ala Val Asp Ala Ala Ile Gly Leu Ala Gln Val Ala Val
 420                425                430

Asp Ala Leu Ala Asp Pro Gln Met Leu Ile Asp Ala Thr Leu Glu Phe
 435                440                445

Thr Asn Ser Gly Gly Met Leu Lys Ala Gly Asp Tyr Leu Ala
 450                455                460

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<210> SEQ ID NO 20
<211> LENGTH: 319
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum

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<400> SEQUENCE: 20

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```

Met Thr Ser Gln Pro Gln Val Arg His Phe Leu Ala Asp Asp Asp Leu
 1                5                10                15

Thr Pro Ala Glu Gln Ala Glu Val Leu Thr Leu Ala Ala Lys Leu Lys
 20                25                30

Ala Ala Pro Phe Ser Glu Arg Pro Leu Glu Gly Pro Lys Ser Val Ala
 35                40                45

Val Leu Phe Asp Lys Thr Ser Thr Arg Thr Arg Phe Ser Phe Asp Ala
 50                55                60

Gly Ile Ala His Leu Gly Gly His Ala Ile Val Val Asp Ser Gly Ser
 65                70                75                80

Ser Gln Met Gly Lys Gly Glu Ser Leu Gln Asp Thr Ala Ala Val Leu
 85                90                95

Ser Arg Tyr Val Glu Ala Ile Val Trp Arg Thr Tyr Ala His Ser Asn
 100               105               110

Phe His Ala Met Ala Glu Thr Ser Thr Val Pro Leu Val Asn Ser Leu
 115               120               125

Ser Asp Asp Leu His Pro Cys Gln Ile Leu Ala Asp Leu Gln Thr Ile
 130               135               140

Val Glu Asn Leu Ser Pro Glu Glu Gly Pro Ala Gly Leu Lys Gly Lys
 145               150               155               160

Lys Ala Val Tyr Leu Gly Asp Gly Asp Asn Asn Met Ala Asn Ser Tyr
 165               170               175

Met Ile Gly Phe Ala Thr Ala Gly Met Asp Ile Ser Ile Ile Ala Pro
 180               185               190

Glu Gly Phe Gln Pro Arg Ala Glu Phe Val Glu Arg Ala Glu Lys Arg

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195					200					205					
Gly	Gln	Glu	Thr	Gly	Ala	Lys	Val	Val	Val	Thr	Asp	Ser	Leu	Asp	Glu
210						215					220				
Val	Ala	Gly	Ala	Asp	Val	Val	Ile	Thr	Asp	Thr	Trp	Val	Ser	Met	Gly
225					230					235					240
Met	Glu	Asn	Asp	Gly	Ile	Asp	Arg	Thr	Thr	Pro	Phe	Val	Pro	Tyr	Gln
				245					250					255	
Val	Asn	Asp	Glu	Val	Met	Ala	Lys	Ala	Asn	Asp	Gly	Ala	Ile	Phe	Leu
			260					265					270		
His	Cys	Leu	Pro	Ala	Tyr	Arg	Gly	Lys	Glu	Val	Ala	Ala	Ser	Val	Ile
		275					280					285			
Asp	Gly	Pro	Ala	Ser	Lys	Val	Phe	Asp	Glu	Ala	Glu	Asn	Arg	Leu	His
	290					295					300				
Ala	Gln	Lys	Ala	Leu	Leu	Val	Trp	Leu	Leu	Ala	Asn	Gln	Pro	Arg	
305					310					315					

<210> SEQ ID NO 21

<211> LENGTH: 533

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 21

Met	Ile	Leu	Gly	Val	Pro	Ile	Gln	Tyr	Leu	Leu	Tyr	Ser	Leu	Trp	Asn
1				5					10					15	
Trp	Ile	Val	Asp	Thr	Gly	Phe	Asp	Val	Ala	Ile	Ile	Leu	Val	Leu	Ala
			20					25					30		
Phe	Leu	Ile	Pro	Arg	Ile	Gly	Arg	Leu	Ala	Met	Arg	Ile	Ile	Lys	Arg
	35					40					45				
Arg	Val	Glu	Ser	Ala	Ala	Asp	Ala	Asp	Thr	Thr	Lys	Asn	Gln	Leu	Ala
	50					55					60				
Phe	Ala	Gly	Val	Gly	Val	Tyr	Ile	Ala	Gln	Ile	Val	Ala	Phe	Phe	Met
65					70				75						80
Leu	Ala	Val	Ser	Ala	Met	Gln	Ala	Phe	Gly	Phe	Ser	Leu	Ala	Gly	Ala
			85						90					95	
Ala	Ile	Pro	Ala	Thr	Ile	Ala	Ser	Ala	Ala	Ile	Gly	Leu	Gly	Ala	Gln
		100						105					110		
Ser	Ile	Val	Ala	Asp	Phe	Leu	Ala	Gly	Phe	Phe	Ile	Leu	Thr	Glu	Lys
		115				120					125				
Gln	Phe	Gly	Val	Gly	Asp	Trp	Val	Arg	Phe	Glu	Gly	Asn	Gly	Ile	Val
	130				135						140				
Val	Glu	Gly	Thr	Val	Ile	Glu	Ile	Thr	Met	Arg	Ala	Thr	Lys	Ile	Arg
145				150					155					160	
Thr	Ile	Ala	Gln	Glu	Thr	Val	Ile	Ile	Pro	Asn	Ser	Thr	Ala	Lys	Val
			165					170						175	
Cys	Ile	Asn	Asn	Ser	Asn	Asn	Trp	Ser	Arg	Ala	Val	Val	Val	Ile	Pro
		180						185					190		
Ile	Pro	Met	Leu	Gly	Ser	Glu	Asn	Ile	Thr	Asp	Val	Ile	Ala	Arg	Ser
		195					200					205			
Glu	Ala	Ala	Thr	Arg	Arg	Ala	Leu	Gly	Gln	Glu	Lys	Ile	Ala	Pro	Glu
	210					215					220				
Ile	Leu	Gly	Glu	Leu	Asp	Val	His	Pro	Ala	Thr	Glu	Val	Thr	Pro	Pro
225				230					235					240	
Thr	Val	Val	Gly	Met	Pro	Trp	Met	Val	Thr	Met	Arg	Phe	Leu	Val	Gln
			245				250							255	

[illegible]

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<210> SEQ ID NO 22
<211> LENGTH: 711
<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum
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<400> SEQUENCE: 22

Met	Lys	Ser	Met	Asn	Ile	Ala	Ala	Ser	Ser	Glu	Leu	Val	Ser	Arg	Leu
1				5					10					15	
Ser	Ser	His	Arg	Arg	Val	Val	Ala	Leu	Gly	Asp	Thr	Asp	Phe	Thr	Asp
			20					25					30		
Val	Ala	Ala	Val	Val	Ile	Thr	Ala	Ala	Asp	Ser	Arg	Ser	Gly	Ile	Leu
		35					40					45			
Ala	Leu	Leu	Lys	Arg	Thr	Gly	Phe	His	Leu	Pro	Val	Phe	Leu	Tyr	Ser
	50					55					60				
Glu	His	Ala	Val	Glu	Leu	Pro	Ala	Gly	Val	Thr	Ala	Val	Ile	Asn	Gly
65					70					75				80	
Asn	Glu	Gln	Gln	Trp	Leu	Glu	Leu	Glu	Ser	Ala	Ala	Cys	Gln	Tyr	Glu
				85					90				95		

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Glu	Asn	Leu	Leu	Pro	Pro	Phe	Tyr	Asp	Thr	Leu	Thr	Gln	Tyr	Val	Glu
		100						105					110		
Met	Gly	Asn	Ser	Thr	Phe	Ala	Cys	Pro	Gly	His	Gln	His	Gly	Ala	Phe
		115					120				125				
Phe	Lys	Lys	His	Pro	Ala	Gly	Arg	His	Phe	Tyr	Asp	Phe	Phe	Gly	Glu
	130					135					140				
Asn	Val	Phe	Arg	Ala	Asp	Met	Cys	Asn	Ala	Asp	Val	Lys	Leu	Gly	Asp
145				150						155					160
Leu	Leu	Ile	His	Glu	Gly	Ser	Ala	Lys	Asp	Ala	Gln	Lys	Phe	Ala	Ala
			165						170					175	
Lys	Val	Phe	His	Ala	Asp	Lys	Thr	Tyr	Phe	Val	Leu	Asn	Gly	Thr	Ser
		180						185					190		
Ala	Ala	Asn	Lys	Val	Val	Thr	Asn	Ala	Leu	Leu	Thr	Arg	Gly	Asp	Leu
		195					200					205			
Val	Leu	Phe	Asp	Arg	Asn	Asn	His	Lys	Ser	Asn	His	His	Gly	Ala	Leu
	210					215					220				
Ile	Gln	Ala	Gly	Ala	Thr	Pro	Val	Tyr	Leu	Glu	Ala	Ser	Arg	Asn	Pro
225					230					235					240
Phe	Gly	Phe	Ile	Gly	Gly	Ile	Asp	Ala	His	Cys	Phe	Asn	Glu	Glu	Tyr
			245					250						255	
Leu	Arg	Gln	Gln	Ile	Arg	Asp	Val	Ala	Pro	Glu	Lys	Ala	Asp	Leu	Pro
		260					265						270		
Arg	Pro	Tyr	Arg	Leu	Ala	Ile	Ile	Gln	Leu	Gly	Thr	Tyr	Asp	Gly	Thr
		275				280						285			
Val	Tyr	Asn	Ala	Arg	Gln	Val	Ile	Asp	Thr	Val	Gly	His	Leu	Cys	Asp
	290					295					300				
Tyr	Ile	Leu	Phe	Asp	Ser	Ala	Trp	Val	Gly	Tyr	Glu	Gln	Phe	Ile	Pro
305					310					315					320
Met	Met	Ala	Asp	Ser	Ser	Pro	Leu	Leu	Leu	Glu	Leu	Asn	Glu	Asn	Asp
			325					330					335		
Pro	Gly	Ile	Phe	Val	Thr	Gln	Ser	Val	His	Lys	Gln	Gln	Ala	Gly	Phe
		340						345					350		
Ser	Gln	Thr	Ser	Gln	Ile	His	Lys	Lys	Asp	Asn	His	Ile	Arg	Gly	Gln
		355				360						365			
Ala	Arg	Phe	Cys	Pro	His	Lys	Arg	Leu	Asn	Asn	Ala	Phe	Met	Leu	His
	370					375					380				
Ala	Ser	Thr	Ser	Pro	Phe	Tyr	Pro	Leu	Phe	Ala	Ala	Leu	Asp	Val	Asn
385					390					395					400
Ala	Lys	Ile	His	Glu	Gly	Glu	Ser	Gly	Arg	Arg	Leu	Trp	Ala	Glu	Cys
		405						410					415		
Val	Glu	Ile	Gly	Ile	Glu	Ala	Arg	Lys	Ala	Ile	Leu	Ala	Arg	Cys	Lys
		420						425					430		
Leu	Phe	Arg	Pro	Phe	Ile	Pro	Pro	Val	Val	Asp	Gly	Lys	Leu	Trp	Gln
		435				440						445			
Asp	Tyr	Pro	Thr	Ser	Val	Leu	Ala	Ser	Asp	Arg	Arg	Phe	Phe	Ser	Phe
	450					455						460			
Glu	Pro	Gly	Ala	Lys	Trp	His	Gly	Phe	Glu	Gly	Tyr	Ala	Ala	Asp	Gln
465					470					475					480
Tyr	Phe	Val	Asp	Pro	Cys	Lys	Leu	Leu	Leu	Thr	Thr	Pro	Gly	Ile	Asp
			485					490						495	
Ala	Glu	Thr	Gly	Glu	Tyr	Ser	Asp	Phe	Gly	Val	Pro	Ala	Thr	Ile	Leu
		500						505						510	

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Ala His Tyr Leu Arg Glu Asn Gly Ile Val Pro Glu Lys Cys Asp Leu
 515 520 525
 Asn Ser Ile Leu Phe Leu Leu Thr Pro Ala Glu Ser His Glu Lys Leu
 530 535 540
 Ala Gln Leu Val Ala Met Leu Ala Gln Phe Glu Gln His Ile Glu Asp
 545 550 555 560
 Asp Ser Pro Leu Val Glu Val Leu Pro Ser Val Tyr Asn Lys Tyr Pro
 565 570 575
 Val Arg Tyr Arg Asp Tyr Thr Leu Arg Gln Leu Cys Gln Glu Met His
 580 585 590
 Asp Leu Tyr Val Ser Phe Asp Val Lys Asp Leu Gln Lys Ala Met Phe
 595 600 605
 Arg Gln Gln Ser Phe Pro Ser Val Val Met Asn Pro Gln Asp Ala His
 610 615 620
 Ser Ala Tyr Ile Arg Gly Asp Val Glu Leu Val Arg Ile Arg Asp Ala
 625 630 635 640
 Glu Gly Arg Ile Ala Ala Glu Gly Ala Leu Pro Tyr Pro Pro Gly Val
 645 650 655
 Leu Cys Val Val Pro Gly Glu Val Trp Gly Gly Ala Val Gln Arg Tyr
 660 665 670
 Phe Leu Ala Leu Glu Glu Gly Val Asn Leu Leu Pro Gly Phe Ser Pro
 675 680 685
 Glu Leu Gln Gly Val Tyr Ser Glu Thr Asp Ala Asp Gly Val Lys Arg
 690 695 700
 Leu Tyr Gly Tyr Val Leu Lys
 705 710

<210> SEQ ID NO 23
 <211> LENGTH: 357
 <212> TYPE: PRT
 <213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 23

Met Ile Met His Asn Val Tyr Gly Val Thr Met Thr Ile Lys Val Ala
 1 5 10 15
 Ile Ala Gly Ala Ser Gly Tyr Ala Gly Gly Glu Ile Leu Arg Leu Leu
 20 25 30
 Leu Gly His Pro Ala Tyr Ala Ser Gly Glu Leu Glu Ile Gly Ala Leu
 35 40 45
 Thr Ala Ala Ser Thr Ala Gly Ser Thr Leu Gly Glu Leu Met Pro His
 50 55 60
 Ile Pro Gln Leu Ala Asp Arg Val Ile Gln Asp Thr Thr Ala Glu Thr
 65 70 75 80
 Leu Ala Gly His Asp Val Val Phe Leu Gly Leu Pro His Gly Phe Ser
 85 90 95
 Ala Glu Ile Ala Leu Gln Leu Gly Pro Asp Val Thr Val Ile Asp Cys
 100 105 110
 Ala Ala Asp Phe Arg Leu Gln Asn Ala Ala Asp Trp Glu Lys Phe Tyr
 115 120 125
 Gly Ser Glu His Gln Gly Thr Trp Pro Tyr Gly Ile Pro Glu Met Pro
 130 135 140
 Gly His Arg Glu Ala Leu Arg Gly Ala Lys Arg Val Ala Val Pro Gly
 145 150 155 160
 Cys Phe Pro Thr Gly Ala Thr Leu Ala Leu Leu Pro Ala Val Gln Ala
 165 170 175

-continued

Gly Leu Ile Glu Pro Asp Val Ser Val Val Ser Ile Thr Gly Val Ser
 180 185 190
 Gly Ala Gly Lys Lys Ala Ser Val Ala Leu Leu Gly Ser Glu Thr Met
 195 200 205
 Gly Ser Leu Lys Ala Tyr Asn Thr Ser Gly Lys His Arg His Thr Pro
 210 215 220
 Glu Ile Ala Gln Asn Leu Gly Glu Val Ser Asp Lys Pro Val Lys Val
 225 230 235 240
 Ser Phe Thr Pro Val Leu Ala Pro Leu Pro Arg Gly Ile Leu Thr Thr
 245 250 255
 Ala Thr Ala Pro Leu Lys Glu Gly Val Thr Ala Glu Gln Ala Arg Ala
 260 265 270
 Val Tyr Glu Glu Phe Tyr Ala Gln Glu Thr Phe Val His Val Leu Pro
 275 280 285
 Glu Gly Ala Gln Pro Gln Thr Gln Ala Val Leu Gly Ser Asn Met Cys
 290 295 300
 His Val Gln Val Glu Ile Asp Glu Glu Ala Gly Lys Val Leu Val Thr
 305 310 315 320
 Ser Ala Ile Asp Asn Leu Thr Lys Gly Thr Ala Gly Ala Ala Val Gln
 325 330 335
 Cys Met Asn Leu Ser Val Gly Phe Asp Glu Ala Ala Gly Leu Pro Gln
 340 345 350
 Val Gly Val Ala Pro
 355

<210> SEQ ID NO 24

<211> LENGTH: 388

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 24

Met Ala Glu Lys Gly Ile Thr Ala Pro Lys Gly Phe Val Ala Ser Ala
 1 5 10 15
 Thr Thr Ala Gly Ile Lys Ala Ser Gly Asn Pro Asp Met Ala Leu Val
 20 25 30
 Val Asn Gln Gly Pro Glu Phe Ser Ala Ala Ala Val Phe Thr Arg Asn
 35 40 45
 Arg Val Phe Ala Ala Pro Val Lys Val Ser Arg Glu Asn Val Ala Asp
 50 55 60
 Gly Gln Ile Arg Ala Val Leu Tyr Asn Ala Gly Asn Ala Asn Ala Cys
 65 70 75 80
 Asn Gly Leu Gln Gly Glu Lys Asp Ala Arg Glu Ser Val Ser His Leu
 85 90 95
 Ala Gln Asn Leu Gly Leu Glu Asp Ser Asp Ile Gly Val Cys Ser Thr
 100 105 110
 Gly Leu Ile Gly Glu Leu Leu Pro Met Asp Lys Leu Asn Ala Gly Ile
 115 120 125
 Asp Gln Leu Thr Ala Glu Gly Ala Leu Gly Asp Asn Gly Ala Ala Ala
 130 135 140
 Ala Lys Ala Ile Met Thr Thr Asp Thr Val Asp Lys Glu Thr Val Val
 145 150 155 160
 Phe Ala Asp Gly Trp Thr Val Gly Gly Met Gly Lys Gly Val Gly Met
 165 170 175
 Met Ala Pro Ser Leu Ala Thr Met Leu Val Cys Leu Thr Thr Asp Ala

-continued

180							185					190				
Ser	Val	Thr	Gln	Glu	Met	Ala	Gln	Ile	Ala	Leu	Ala	Asn	Ala	Thr	Ala	
		195					200					205				
Val	Thr	Phe	Asp	Thr	Leu	Asp	Ile	Asp	Gly	Ser	Thr	Ser	Thr	Asn	Asp	
	210					215					220					
Thr	Val	Phe	Leu	Leu	Ala	Ser	Gly	Ala	Ser	Gly	Ile	Thr	Pro	Thr	Gln	
	225				230					235					240	
Asp	Glu	Leu	Asn	Asp	Ala	Val	Tyr	Ala	Ala	Cys	Ser	Asp	Ile	Ala	Ala	
				245					250					255		
Lys	Leu	Gln	Ala	Asp	Ala	Glu	Gly	Val	Thr	Lys	Arg	Val	Ala	Val	Thr	
			260					265					270			
Val	Val	Gly	Thr	Thr	Asn	Asn	Glu	Gln	Ala	Ile	Asn	Ala	Ala	Arg	Thr	
		275					280					285				
Val	Ala	Arg	Asp	Asn	Leu	Phe	Lys	Cys	Ala	Met	Phe	Gly	Ser	Asp	Pro	
	290					295					300					
Asn	Trp	Gly	Arg	Val	Leu	Ala	Ala	Val	Gly	Met	Ala	Asp	Ala	Asp	Met	
	305				310					315					320	
Glu	Pro	Glu	Lys	Ile	Ser	Val	Phe	Phe	Asn	Gly	Gln	Ala	Val	Cys	Leu	
				325					330					335		
Asp	Ser	Thr	Gly	Ala	Pro	Gly	Ala	Arg	Glu	Val	Asp	Leu	Ser	Gly	Ala	
			340					345					350			
Asp	Ile	Asp	Val	Arg	Ile	Asp	Leu	Gly	Thr	Ser	Gly	Glu	Gly	Gln	Ala	
		355					360					365				
Thr	Val	Arg	Thr	Thr	Asp	Leu	Ser	Phe	Ser	Tyr	Val	Glu	Ile	Asn	Ser	
	370					375					380					
Ala	Tyr	Ser	Ser													
	385															
<210> SEQ ID NO 25																
<211> LENGTH: 317																
<212> TYPE: PRT																
<213> ORGANISM: Corynebacterium glutamicum																
<400> SEQUENCE: 25																
Met	Asn	Asp	Leu	Ile	Lys	Asp	Leu	Gly	Ser	Glu	Val	Arg	Ala	Asn	Val	
1				5					10					15		
Leu	Ala	Glu	Ala	Leu	Pro	Trp	Leu	Gln	His	Phe	Arg	Asp	Lys	Ile	Val	
		20						25					30			
Val	Val	Lys	Tyr	Gly	Gly	Asn	Ala	Met	Val	Asp	Asp	Asp	Leu	Lys	Ala	
		35					40					45				
Ala	Phe	Ala	Ala	Asp	Met	Val	Phe	Leu	Arg	Thr	Val	Gly	Ala	Lys	Pro	
	50					55					60					
Val	Val	Val	His	Gly	Gly	Gly	Pro	Gln	Ile	Ser	Glu	Met	Leu	Asn	Arg	
	65			70						75				80		
Val	Gly	Leu	Gln	Gly	Glu	Phe	Lys	Gly	Gly	Phe	Arg	Val	Thr	Thr	Pro	
			85					90						95		
Glu	Val	Met	Asp	Ile	Val	Arg	Met	Val	Leu	Phe	Gly	Gln	Val	Gly	Arg	
		100						105					110			
Asp	Leu	Val	Gly	Leu	Ile	Asn	Ser	His	Gly	Pro	Tyr	Ala	Val	Gly	Thr	
		115					120					125				
Ser	Gly	Glu	Asp	Ala	Gly	Leu	Phe	Thr	Ala	Gln	Lys	Arg	Met	Val	Asn	
	130					135						140				
Ile	Asp	Gly	Val	Pro	Thr	Asp	Ile	Gly	Leu	Val	Gly	Asp	Ile	Ile	Asn	
	145				150					155					160	

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Val Asp Ala Ser Ser Leu Met Asp Ile Ile Glu Ala Gly Arg Ile Pro
      165                      170                      175

Val Val Ser Thr Ile Ala Pro Gly Glu Asp Gly Gln Ile Tyr Asn Ile
      180                      185                      190

Asn Ala Asp Thr Ala Ala Gly Ala Leu Ala Ala Ala Ile Gly Ala Glu
      195                      200                      205

Arg Leu Leu Val Leu Thr Asn Val Glu Gly Leu Tyr Thr Asp Trp Pro
      210                      215                      220

Asp Lys Ser Ser Leu Val Ser Lys Ile Lys Ala Thr Glu Leu Glu Ala
      225                      230                      235                      240

Ile Leu Pro Gly Leu Asp Ser Gly Met Ile Pro Lys Met Glu Ser Cys
      245                      250                      255

Leu Asn Ala Val Arg Gly Gly Val Ser Ala Ala His Val Ile Asp Gly
      260                      265                      270

Arg Ile Ala His Ser Val Leu Leu Glu Leu Leu Thr Met Gly Gly Ile
      275                      280                      285

Gly Thr Met Val Leu Pro Asp Val Phe Asp Arg Glu Asn Tyr Pro Glu
      290                      295                      300

Gly Thr Val Phe Arg Lys Asp Asp Lys Asp Gly Glu Leu
      305                      310                      315

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<210> SEQ ID NO 26

<211> LENGTH: 391

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 26

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Met Ser Thr Leu Glu Thr Trp Pro Gln Val Ile Ile Asn Thr Tyr Gly
1      5      10      15

Thr Pro Pro Val Glu Leu Val Ser Gly Lys Gly Ala Thr Val Thr Asp
      20      25      30

Asp Gln Gly Asn Val Tyr Ile Asp Leu Leu Ala Gly Ile Ala Val Asn
      35      40      45

Ala Leu Gly His Ala His Pro Ala Ile Ile Glu Ala Val Thr Asn Gln
      50      55      60

Ile Gly Gln Leu Gly His Val Ser Asn Leu Phe Ala Ser Arg Pro Val
      65      70      75      80

Val Glu Val Ala Glu Glu Leu Ile Lys Arg Phe Ser Leu Asp Asp Ala
      85      90      95

Thr Leu Ala Ala Gln Thr Arg Val Phe Phe Cys Asn Ser Gly Ala Glu
      100     105     110

Ala Asn Glu Ala Ala Phe Lys Ile Ala Arg Leu Thr Gly Arg Ser Arg
      115     120     125

Ile Leu Ala Ala Val His Gly Phe His Gly Arg Thr Met Gly Ser Leu
      130     135     140

Ala Leu Thr Gly Gln Pro Asp Lys Arg Glu Ala Phe Leu Pro Met Pro
      145     150     155     160

Ser Gly Val Glu Phe Tyr Pro Tyr Gly Asp Thr Asp Tyr Leu Arg Lys
      165     170     175

Met Val Glu Thr Asn Pro Thr Asp Val Ala Ala Ile Phe Leu Glu Pro
      180     185     190

Ile Gln Gly Glu Thr Gly Val Val Pro Ala Pro Glu Gly Phe Leu Lys
      195     200     205

Ala Val Arg Glu Leu Cys Asp Glu Tyr Gly Ile Leu Met Ile Thr Asp
      210     215     220

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Glu Val Gln Thr Gly Val Gly Arg Thr Gly Asp Phe Phe Ala His Gln
 225 230 235 240
 His Asp Gly Val Val Pro Asp Val Val Thr Met Ala Lys Gly Leu Gly
 245 250 255
 Gly Gly Leu Pro Ile Gly Ala Cys Leu Ala Thr Gly Arg Ala Ala Glu
 260 265 270
 Leu Met Thr Pro Gly Lys His Gly Thr Thr Phe Gly Gly Asn Pro Val
 275 280 285
 Ala Cys Ala Ala Ala Lys Ala Val Leu Ser Val Val Asp Asp Ala Phe
 290 295 300
 Cys Ala Glu Val Ala Arg Lys Gly Glu Leu Phe Lys Glu Leu Leu Ala
 305 310 315 320
 Lys Val Asp Gly Val Val Asp Val Arg Gly Arg Gly Leu Met Leu Gly
 325 330 335
 Val Val Leu Glu Arg Asp Val Ala Lys Gln Ala Val Leu Asp Gly Phe
 340 345 350
 Lys His Gly Val Ile Leu Asn Ala Pro Ala Asp Asn Ile Ile Arg Leu
 355 360 365
 Thr Pro Pro Leu Val Ile Thr Asp Glu Glu Ile Ala Asp Ala Val Lys
 370 375 380
 Ala Ile Ala Glu Thr Ile Ala
 385 390

<210> SEQ ID NO 27

<211> LENGTH: 538

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 27

Met Ser Ser Thr Thr Ser Lys Thr Ser Glu Arg Gln Gln Pro Asp Ala
 1 5 10 15
 Pro Thr Ser Lys Leu Ser Lys Trp Ser Asp Lys Phe Leu Asn Gly Val
 20 25 30
 Glu Thr Leu Gly Asn Lys Leu Pro Thr Pro Phe Thr Leu Phe Leu Ile
 35 40 45
 Leu Phe Leu Ile Thr Ala Leu Ala Ser Ser Ile Met Ala Trp Met Asn
 50 55 60
 Val Ser Val Ile Val Pro Gly Ser Asp Glu Glu Leu Phe Val Lys Gly
 65 70 75 80
 Leu Phe Thr Gly Glu Gly Leu Thr Trp Leu Thr Thr Asn Leu Gly Ala
 85 90 95
 Asn Tyr Ile Gly Phe Pro Pro Leu Leu Thr Val Leu Pro Ile Leu Leu
 100 105 110
 Ala Val Gly Val Ala Glu Arg Ser Gly Met Leu Ala Ala Leu Ile Arg
 115 120 125
 Lys Leu Phe Gly Ser Ala Lys Lys Ile Val Leu Pro Tyr Ala Val Gly
 130 135 140
 Val Ile Gly Val Thr Ala Ser Ile Met Ala Asp Ala Ala Phe Val Val
 145 150 155 160
 Val Pro Pro Leu Ala Ala Met Val Phe Lys Ala Ala Gly Arg His Pro
 165 170 175
 Val Ala Gly Leu Leu Gly Ser Phe Ala Ala Val Gly Ala Gly Tyr Ser
 180 185 190
 Thr Ala Ile Val Pro Thr Ser Leu Asp Ala Leu Phe Ala Gly Ile Thr

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195					200					205					
Asn	Ala	Val	Met	Glu	Thr	Leu	Pro	Gly	Ile	Ala	Thr	Thr	Glu	Val	Asn
210						215					220				
Pro	Val	Ser	Asn	Tyr	Tyr	Phe	Asn	Ile	Ala	Ser	Ser	Ile	Val	Leu	Gly
225					230					235				240	
Leu	Leu	Cys	Gly	Phe	Leu	Ile	Asp	Lys	Val	Leu	Glu	Pro	Arg	Met	Trp
				245					250					255	
Arg	Gln	Lys	Ile	Ala	Thr	Glu	Tyr	Ala	Glu	Ser	Ile	Glu	Pro	Thr	Ser
			260					265					270		
Ala	Ala	Asp	Asp	Glu	Glu	Ile	Ser	Ala	Thr	Leu	Thr	Ala	Gln	Glu	Asn
		275					280					285			
Arg	Ala	Leu	Thr	Ile	Ser	Met	Trp	Thr	Thr	Leu	Ala	Thr	Ala	Ile	Ile
	290					295				300					
Val	Leu	Val	Val	Val	Leu	Ile	Pro	Gly	Ser	Pro	Trp	Arg	Asn	Glu	Asp
305					310					315				320	
Gly	Gly	Phe	Leu	Pro	Thr	Ser	Pro	Leu	Leu	Ser	Ser	Val	Val	Phe	Ile
				325				330						335	
Val	Phe	Leu	Phe	Phe	Met	Val	Met	Gly	Leu	Ala	Tyr	Gly	Met	Val	Val
			340					345					350		
Gly	Thr	Ile	Lys	Asn	Met	Asp	Asp	Val	Val	Asn	Met	Met	Gly	Glu	Ala
			355				360					365			
Ile	Lys	Asp	Met	Ile	Gly	Phe	Leu	Val	Leu	Ala	Phe	Ile	Leu	Gly	Gln
	370					375					380				
Phe	Val	Ala	Leu	Phe	Asn	Trp	Thr	Gly	Ile	Gly	Thr	Trp	Thr	Ala	Val
385					390					395				400	
Gln	Gly	Ala	Ala	Gly	Leu	Glu	Ala	Ile	Gly	Leu	Thr	Gly	Phe	Pro	Ala
			405					410						415	
Ile	Ile	Ala	Phe	Ile	Ile	Leu	Ala	Ser	Cys	Leu	Asn	Leu	Leu	Ile	Ile
			420					425					430		
Ser	Gly	Ser	Ala	Met	Trp	Thr	Leu	Met	Ala	Ala	Val	Phe	Val	Pro	Met
	435						440					445			
Phe	Ala	Leu	Leu	Gly	Tyr	Glu	Pro	Ser	Phe	Ile	Gln	Ala	Ala	Phe	Arg
	450					455					460				
Val	Gly	Asp	Ser	Ala	Thr	Gln	Val	Ile	Thr	Pro	Leu	Asn	Pro	Tyr	Met
465				470						475				480	
Ile	Val	Ile	Leu	Gly	Leu	Leu	Arg	Arg	Tyr	Glu	Pro	Asp	Ala	Gly	Leu
			485					490						495	
Gly	Thr	Leu	Met	Ser	Arg	Leu	Ile	Pro	Phe	Val	Ile	Pro	Phe	Trp	Leu
		500						505					510		
Ala	Trp	Ala	Thr	Leu	Leu	Ala	Ile	Trp	Phe	Tyr	Ala	Asp	Leu	Pro	Leu
	515						520					525			
Gly	Pro	Gly	Ser	Ala	Ile	Phe	Leu	Glu	Gly						
	530					535									

<210> SEQ ID NO 28

<211> LENGTH: 102

<212> TYPE: PRT

<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 28

Met	Trp	Thr	Leu	Met	Ala	Ala	Val	Phe	Val	Pro	Met	Phe	Ala	Leu	Leu
1				5				10						15	

Gly	Tyr	Glu	Pro	Ser	Phe	Ile	Gln	Ala	Ala	Phe	Arg	Val	Gly	Asp	Ser
	20						25						30		

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Ala Thr Gln Val Ile Thr Pro Leu Asn Pro Tyr Met Ile Val Ile Leu
 35 40 45

Gly Leu Leu Arg Arg Tyr Glu Pro Asp Ala Gly Leu Gly Thr Leu Met
 50 55 60

Ser Arg Leu Ile Pro Phe Val Ile Pro Phe Trp Leu Ala Trp Ala Thr
 65 70 75 80

Leu Leu Ala Ile Trp Phe Tyr Ala Asp Leu Pro Leu Gly Pro Gly Ser
 85 90 95

Ala Ile Phe Leu Glu Gly
 100

<210> SEQ ID NO 29
 <211> LENGTH: 122
 <212> TYPE: PRT
 <213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 29

Val Asp Glu Arg Ser Arg Phe Ala Arg Ser Val Phe Pro Asp Gly Glu
 1 5 10 15

Glu Pro Asp Pro Arg Phe Thr Leu Ala Asn Glu Arg Thr Phe Leu Ala
 20 25 30

Trp Thr Arg Thr Ser Leu Ala Phe Leu Ala Gly Gly Ile Ala Phe Glu
 35 40 45

Ala Phe Gln Ile Ser Gly Leu Ser Asp Thr Val Arg Thr Thr Ile Ala
 50 55 60

Val Phe Ile Ile Ala Val Gly Met Ile Ile Ala Ala Gly Ala Ala Val
 65 70 75 80

Arg Trp Met Asn Val Glu Arg Ala Met Arg Lys Gln Lys Pro Leu Pro
 85 90 95

Val Pro Ala Ile Ile Pro Phe Leu Ser Ile Ala Ala Leu Val Ala Ser
 100 105 110

Ala Ala Val Leu Val Leu Ile Ile Val Gln
 115 120

<210> SEQ ID NO 30
 <211> LENGTH: 105
 <212> TYPE: PRT
 <213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 30

Met Arg Ile His Glu Asp Pro Gly Leu Gln Pro Glu Arg Thr Val Leu
 1 5 10 15

Ala Trp Asn Arg Thr Thr Val Ser Leu Ala Val Cys Ser Ala Ile Leu
 20 25 30

Leu Arg Trp Thr Asn Phe Tyr Gly Ile Phe Ala Leu Leu Pro Val Val
 35 40 45

Ile Leu Ser Gly Met Ala Ile Phe Ile Leu Phe Thr Gln Arg Val Arg
 50 55 60

Tyr Glu Arg Gln Ala Ile Gly Leu Ala Asp Asn Lys Leu Pro Pro Asn
 65 70 75 80

Ile Val Gly Val Val Ser Leu Thr Val Thr Leu Leu Ala Phe Gly Ala
 85 90 95

Ala Gly Ile Val Phe Val Phe Ile Asp
 100 105

<210> SEQ ID NO 31
 <211> LENGTH: 265

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<212> TYPE: PRT
<213> ORGANISM: Corynebacterium glutamicum

<400> SEQUENCE: 31
Val Ala Gly Asp Lys Leu Trp Leu Cys Asp Val His Phe Pro Val Ala
1      5      10      15
Arg Cys Trp Glu Thr Thr Thr Gly Arg Tyr Leu Gly Gln Thr Leu Val
20      25      30
Pro Ala Pro Leu Arg Asp Arg Ser Tyr Val Leu Glu Leu His Ser Asp
35      40      45
Gln Gln Leu Gly Ala Val Ala Ala Ser Gly Lys Ser Gly Trp Ile Leu
50      55      60
Thr Pro Gly Gln Ala Val Ala Thr Lys Ala Pro Asp Trp Thr Pro Pro
65      70      75      80
Thr Arg Ala Thr Asp Leu Pro Gln Val Pro Ser Pro Trp Glu Ile Val
85      90      95
Ala Val Arg Gly Gln Gly Leu Phe Glu Leu Gln Val Glu Thr Ser Arg
100     105     110
Arg Thr Ala Leu Gly Arg Val Asn Ala Thr Gly Gly Val Asp Ile Gly
115     120     125
Glu Leu Pro Pro Asn Gly Tyr Thr Ile Ser Ser Val Val Gln Ile Gly
130     135     140
Asp Glu Tyr Ile Val Glu Arg Trp Val Glu Glu Tyr Arg Leu Asn Ser
145     150     155     160
Lys Leu Glu Val Ile Ser Thr Lys Glu Leu Asp Ile Ser Ala Ser Gly
165     170     175
Trp Lys Ser Lys Gly Thr Val Ala Tyr Leu Ser Glu Asp Thr His Ile
180     185     190
Cys Phe Phe Asp Gln Val Ser Gly Ala Glu Leu Pro Ser Leu Gly Ile
195     200     205
Ala Glu Gly His Gln Gly Glu Val Met Ser Ala Thr Ser Ser Glu Ser
210     215     220
Ile Val Leu Ile Tyr Arg Arg Asn Pro Asn Asn Ser Met Ser Ile Val
225     230     235     240
Pro Thr Ser Val Ala Thr Tyr Asp Asn Gly Thr Trp Thr Thr Met Pro
245     250     255
Leu Gln Glu Ala Pro Ala Glu Leu Ser
260     265

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The invention claimed is:

1. A recombinant *Corynebacterium glutamicum* having enhanced ability to produce putrescine, wherein the activity of a protein having an amino acid sequence of SEQ ID NO: 17 or SEQ ID NO: 19 is down regulated or removed, compared to the endogenous activity thereof.

2. The recombinant *Corynebacterium glutamicum* according to claim 1, wherein ornithine decarboxylase activity is introduced into the recombinant *Corynebacterium glutamicum*.

3. The recombinant *Corynebacterium glutamicum* according to claim 2, wherein the ornithine decarboxylase has the amino acid sequence of SEQ ID NO: 22.

4. The recombinant *Corynebacterium glutamicum* according to claim 1, wherein ornithine carbamoyltransferase (ArgF) activity and/or glutamate exporter activity are down regulated compared to the endogenous activity thereof.

5. The recombinant *Corynebacterium glutamicum* according to claim 4, wherein the ArgF has the amino acid sequence of SEQ ID NO: 20, and glutamate exporter has the amino acid sequence of SEQ ID NO: 21.

6. The recombinant *Corynebacterium glutamicum* according to claim 1, wherein one or more activities selected from the group consisting of acetyl gamma glutamyl phosphate reductase (ArgC) activity, acetyl glutamate synthase activity or ornithine acetyltransferase (ArgJ) activity, acetyl glutamate kinase (ArgB) activity, and acetyl ornithine amino transferase (ArgD) activity are further enhanced.

7. The recombinant *Corynebacterium glutamicum* according to claim 6, wherein ArgC, ArgJ, ArgB and ArgD have the amino acid sequences of SEQ ID NOs: 23, 24, 25, and 26, respectively.

8. The recombinant *Corynebacterium glutamicum* according to claim 1, wherein the activity of the protein is down regulated by 1) a partial or whole deletion of a polynucleotide

encoding the protein, 2) a reduction of the polynucleotide expression, 3) a modification of the polynucleotide sequence on chromosome to down regulate the activity of the protein or 4) a combination thereof.

9. A method for producing putrescine, comprising cultur- 5
ing a recombinant *Corynebacterium glutamicum* having
enhanced ability to produce putrescine in a cell culture broth,
wherein the activity of a protein having an amino acid
sequence of SEQ ID NO: 17 or SEQ ID NO: 19 is down
regulated or removed in the recombinant *Corynebacterium* 10
glutamicum compared to the endogenous activity thereof;
obtaining the cell culture broth; and isolating putrescine from
the obtained cell culture broth.

* * * * *